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Sequestering agents for uranyl chelation: new calixarene ligands

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1. Introduction

Commonly used as nuclear fuel in fission reactors for civilian purpose, uranium can be introduced into the body in the case of internal contamination or in the event of a nuclear accident by ingestion, inhalation or through wounds. The hexavalent uranyl ion $(UO_2^{2+}, U(VI))$ was found to be the most stable form in vivo¹ and is complexed in the blood by chelating agents such as proteins or carbonates. Distribution of toxic species and retention in target organs such as kidneys, liver or marrow occurs² after chelation, potentially inducing cancer and chemical intoxication, especially in the case of heavy contamination.³

Elimination of toxic species from the body could be achieved by administrating non-toxic chelating agents, which must have high stability constant so that they can displace the natural complexes rapidly formed with components of blood. To lower uranyl concentrations and radiation doses, and subsequently tumour risks, the uranyl/ligand complex formed must also be soluble in physiological fluids and stable in a pH range of 2–9 to be subsequently eliminated from the body by crossing renal or hepatic barrier.

During the past 30 years, several effective uranyl ligands were synthesised, based on different complexing functions. Phosphorus containing molecules, especially bisphosphonates, were found to be very effective uranyl ligands,⁴ but few significant decorporation

ABSTRACT

Synthesis of sulfocatecholamide (CAMS) and hydroxypyridinone (HOPO) calixarene ligands and determination of their binding abilities for the uranyl cation were described. Chelating properties were determined by UV spectrophotometry in aqueous media under various pH conditions and further studied by ¹H NMR analysis of the resonance signals of both aromatics' protons of the chelating groups. Each ligand shows a more or less pronounced affinity for uranium. HOPO calixarenes exhibit significant affinity towards uranyl ion at acidic and neutral pH while CAMS calixarene is more efficient at basic pH. © 2008 Elsevier Ltd. All rights reserved.

work has been reported so far concerning the decorporation efficacy of ethane-1-hydroxy-1,1-bisphosphonate EHBP.⁵ Decorporation with bidentate methylterphthalimide (MeTAM)-based chelating ligands was also studied and appeared not to be suitable for biological decorporation due to their high toxicity.⁶

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Sulfocatechol Tiron proved to be effective for U(VI) complexation in vivo within the physiological pH range,⁷ but a modest successful reduction of acute U(VI) toxicity and reduction of body U(VI) with this ligand was observed. Therefore, multidentate analogues containing sulfocatecholamide (CAMS) or structurally analogous hydroxyl-pyridone (HOPO) units would be effective for in vivo chelation of U(VI).⁸

Uranyl-sequestering agents based on 3-hydroxy-2(1*H*)-pyridinone (3,2-HOPO) and sulfocatecholamide (CAMS) ligands resulted in two low-toxicity ligands 5-LICAM(S) and 5-LIO(Me-3,2-HOPO) (Fig. 1), both efficient chelating agents of circulating U(VI) in the body.^{8b} However, these efficiencies were often observed when the ligands were administrated immediately (5–30 min) after contamination, which shows that the development of new ligands is still of interest.

Recently, we described the synthesis and the evaluation of several 5-CAMS analogues incorporating various diamine skeletons.⁹ The chelating properties towards uranium were studied in aqueous media by UV–vis analysis and NMR spectroscopy and some of these showed pronounced affinity for the target ion (CYCAMS).

Owing to the development of supramolecular chemistry, a number of calixarene-based ligands have been extensively developed for their coordination properties.¹⁰ Moreover, the chemistry of these macrocycles is presently well known and efficient.



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Figure 1. Low-toxicity uranyl complexants.



Figure 2. Calixarenes based uranophiles.

Synthetic modifications can be realised on the lower and/or the upper rim with various functional chelating groups. The easy accessibility and the selective functionalisations at the phenolic hydroxy groups of calix[4]arenes have made this member of the series increasingly attractive for chemists involved in host/guest chemistry.¹¹ Since sulfonato-calix[*n*]arenes (*n*=4,6,8) have been reported¹² as selective uranophiles (Fig. 2), the importance of structural rigidity and pre-organisation of these macrocycles was proved with

the very high stability of the complexes with UO_2^{2+} of the calixarene-based ligand for the uranyl ion relative to other metal ions.¹³ Calix[4]arene systems containing amides,¹⁴ hydroxamates,¹⁵ CMPO¹⁶ or HOPO¹⁷ have also been designed expressively for the selective coordination of uranyl ions.

Surprisingly, to the best of our knowledge, combination of the architecture of calix[*n*]arenes with the chelating behaviour of sulfocatechol amides has not been reported in the literature so far. We present here the synthesis and the chelating properties of new calixarene derivatives containing HOPO or CAMS function.

The first step in this project involves the condensation of protected chelating groups on bis-ethoxyamino calix[4]arenes **3a,b** (Scheme 1). After deprotection of the hydroxyl groups of the chelating units CAM and HOPO, sulfonation afforded the target compounds. The second step concerns the comparative evaluation of the complexation constants with uranyl ions in water with 5-LICAM(S) as a reference, using SCP method developed by Taran and co-workers.^{4a}

2. Results and discussion

2.1. Synthesis

Acid chloride derivatives **1** and **2** were obtained by the reaction of oxalyl chloride with *O*-benzyl catechol¹⁸ and *N*-benzyl HOPO¹⁹ carboxylic acids preliminarily synthesised following the previously described procedures in dichloromoethane with a catalytic amount of DMF in quantitative yield. Bis-catecholamide analogues **4a,b** and **5a,b** were obtained by condensation of the acid chloride derivatives **1** and **2** with calixarenes **3a** and **3b** prepared according to a described procedure²⁰ in the presence of Et₃N (Scheme 1). Deprotection of the hydroxyl groups was achieved using HCl in acetic acid for the HOPO pendant arms to give **6b** and **7b** in 87% and 70% yields, respectively. Hydrogenolysis of the catechol **4a** and **5a** led to **6a** and **7a** in 89% and 98% yields.

Sulfonation in hot sulfuric acid of **6a** or **7a** followed by precipitation in diethylether gave the desired pure 1,3-CalixCAMS **8a** in good yield (Scheme 2). In parallel, sulfonation of 1,3-CalixHOPO **6b** or **7b** failed, leading to partial and total removal of HOPO groups



Scheme 1. Synthesis of 1,3-CalixCAM and CalixHOPO.



Scheme 2. Sulfonation of CAM analogues.

from the calixarene ring. Each component was fully characterised by ¹H NMR, ¹³C NMR and mass spectroscopy. Cone conformation of the macrocycles was confirmed by ¹H NMR that displays two doublets at 3.25–3.48/4.50–4.17 ppm corresponding to the resonance signals of H_{ax} and H_{eq} of the methylene bridges and ¹³C NMR with signals at 31.0–32.1 ppm corresponding to the resonance signals of the methylene bridges.²¹

2.2. Constant stability determination

The complexation behaviour of water-soluble calixarenes 6b, 7b and **8a** towards the uranyl cation was studied by the spectrophotometric method developed by Taran co-workers,^{4a} based on competitive uranium binding by using sulfochlorophenol SCP as a chromogenic chelate. This latter was found highly suitable for a rapid screening of putative uranium ligand library and compared with 5-LICAMS, synthesised as previously described by Raymond and co-workers (Table 1).⁶ Globally, the CAMS macrocycles exhibit high K_{cond} enhancement in basic conditions in accordance with previous findings.²² At pH=7.4, none of the synthesised calixarene displaced SCP/uranyl complexation equilibrium better than the 5-LICAMS with log K_{pH=74}=17.0. Except for the 1,3-CalixCAMS 8a, which was not able to displace more than 20% of the SCP/UO₂ complex,^{4a} 1,3-calixHOPO and 5-LICAMS exhibited similar log K_{cond} close to 11. At pH 9, 8a exhibited a larger complexation efficiency $(\log K_{cond}=20.2)$ towards UO_2^{2+} . As far as we know, such very large stability constant has never been observed with CAMS ligands.

The ability of compound **8a** to complex UO_2^{2+} was also studied by ¹H NMR spectroscopy. Due to their low solubility in D₂O, such studies couldn't be achieved with compounds **6b** and **7b**.

Figure 3 shows the NMR spectra of **8a** with variable amounts of uranyl nitrate hexahydrate in the presence of a binary mixture of D₂O/NaOD at room temperature with sodium nitrate (0.05 mol L⁻¹). After each addition of UO₂(NO₃)₂·6H₂O, the pH of the deuterated solution was adjusted 12 by addition of NaOD.

Resonance signals of the catechol protons H_1 and H_2 of compound **8a** (Fig. 3) in D₂O/NaOD were, respectively, identified as two doublets at 7.62 and 6.97 ppm (*J*=1.9 Hz), while resonance signals of the aromatic protons H_3 and H_4 of the phenolic ring were located as two singlets at 7.42 and 7.68 ppm. Addition of increasing amounts of UO₂(NO₃)₂·6H₂O (Fig. 3) led to appreciable changes

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Table 1

рН	log K _{cond} U-L/(pH	I)	
	5.5	7.4	9.0
5-LICAMS	11.1	17.0	19.4
8a	10	16.3	20.2
6b	11	16	18.1
7b	11.2	15.5	17.9

with downfield shifts of the H_1/H_2 aromatic proton resonances of the catechol moiety and upfield shifts of the H_3/H_4 aromatic proton resonance signals of the calixarene ring.

Thus, upon addition of UO₂(NO₃)₂·6H₂O in a deuterated solution containing **8a**, two new doublets appear, respectively, at 7.72 and 7.13 ppm (Δ_{δ} =0.10 and 0.16 ppm) corresponding to the resonance signals of the catechol protons of the uranyl complexes. Similarly, the changes in the level of the signals of the phenolic protons of the calixarene ring were also indicative due to the proximity of the uranyl cation nearby the altered protons. Thus, two new broad singlets appear, respectively, at 7.28 ppm and 7.54 ppm (Δ_{δ} =-0.14 ppm). Finally, the 1:1 stoichiometry of the uranyl complexes was confirmed by the total disappearance of the four resonance signals of the free ligand without appearance of new resonance signals.

3. Conclusion

The dipodal bis-catecholamide **8a** and bis-HOPO uranophiles **6b**, **7b** were obtained by an efficient synthetic route from diaminocalixarenes **3a,b**. Their binding abilities for UO_2^{2+} were determined by UV spectrophotometry in aqueous media under acidic, neutral and basic pH conditions. Globally, the binding properties were found similar to those observed with 5-LICAMS, a well known ligand that displays in vivo uranyl removal capabilities. At pH=9.0, the CalixCAMS **8a** showed a higher constant of 20.2 never observed with CAMS ligands.

4. Experimental part

4.1. General

All the organic reagents used were pure commercial products from Aldrich, Acros, Fluka, Avocado, Lancaster & Maybridge. Diaminocalixarenes **3a** and **3b** in cone conformation,²⁰ (2,3)diben-zyloxybenzoic acid¹⁸ and 1-hydroxy-6-oxo-1,6-dihydropyridine-2-carboxylic acid (1,2-HOPO)^{19a} were synthesised as previously described.

The solvents were purchased from Carlo Erba, Acros, Pro-Labo, Fulka & Aldrich. Anhydrous solvents came from Acros, anhydrous THF and dry CH₂Cl₂ were distilled. Flash chromatography was carried out on Merck Silica Si60 (40–63 μ m). ¹H, ¹³C NMR spectra were recorded on a Bruker AC-200 (200.13 MHz for ¹H, 50.32 MHz for ¹³C) or AC-300 FT (300.13 MHz for ¹H, 75.46 MHz for ¹³C) spectrometer; δ values are given in parts per million and *J* in hertz. Elemental analyses (C, H, N, S, O, F) were obtained from the Service Central d'Analyse of the CNRS (Solaize). High resolution mass spectra: HR LSIMS (Liquid Secondary Ionisation Mass Spectrometry: Thioglycerol), HR CIMS (Isobutan) and HR EIMS were carried out on a Finnegan MAT 95xL by the UCBL Centre de Spectroscopie de Masse.

4.2. Synthesis of calixarene 4a

Oxalyl chloride (0.8 ml, 9.3 mmol) was added dropwise to a solution of 2.04 g 2,3-bis(benzyloxy)benzoic acid (6.1 mmol) in CH₂Cl₂ (30 ml). After adding a drop of DMF, the mixture was stirred until the end of HCl release. After evaporation of solvents and residual oxalyl chloride, the residue was dissolved in 30 ml of dry CH₂Cl₂ and added dropwise to a solution of 1.53 g of 25,27-(2-aminoethoxy)-26,28-dihydroxycalix[4]arene (3 mmol) and 1.3 ml of triethylamine (9.2 mmol) in 40 ml of dry CH₂Cl₂. After 18 h under stirring, the mixture was washed with HCl 1 N (100 ml), water (2×50 ml), brine (100 ml), then dried over MgSO₄ and evaporated to dryness. The residue was purified by silica gel column chromatography (EtOAc/cyclohexane 1:2) to give **4a** (1.78 g, 52%) as white powder. ¹H NMR (CDCl₃, 300 MHz, 25 °C) δ (ppm): 8.19 (br s, 2H, NH), 7.54 (m, 2H,



Figure 3. ¹H NMR aromatic shifts upon addition of $UO_2(NO_3)_2 - 6H_2O$ in $D_2O+NaOD$ at pH 12 (a₁: 0 equiv; a₂: 0.5 equiv; a₃: 1 equiv) at 298 K (\odot : H₁, \Box : H₂, \diamond : H₃, \triangle : H₄ of the free 1,3-CalixCAMS and \bullet : H₁, \blacksquare : H₂, \diamond : H₃, \triangle : H₄ of 1,3-CalixCAMS/UO₂²⁺).

Ar–H), 7.25–7.37 (m, 20H, Ar–H), 6.97–7.09 (m, 8H, Ar–H), 6.78 (d, J=7.5 Hz, 4H, Ar–H), 6.64 (t, J=4.7 Hz, 4H, Ar–H), 5.07 (s, 4H, O–CH₂–Ar), 4.99 (s, 4H, O–CH₂–Ar), 4.13 (AB d, 4H, 2 J(H,H)=13.3 Hz, Ar–CH₂–Ar ax), 3.90 (t, 4H, J=5.6 Hz, O–CH₂–CH₂–NH), 3.49 (m, 4H, O–CH₂–CH₂–NH), 3.22 (AB d, 4H, 2 J(H,H)=13.3 Hz, Ar–CH₂–Ar eq). ¹³C NMR (CDCl₃, 75 MHz, 25 °C) δ (ppm): 166.8 (C=O), 153.3 (ArC), 152.3 (ArC), 151.7 (ArC), 147.3 (ArC), 137.3 (ArC), 137.1 (ArC), 133.4 (ArC), 129.5 (ArC), 129.4 (ArCH), 129.3 (ArCH), 129.0 (ArCH), 128.9 (ArCH), 128.6 (ArCH), 128.4 (ArC), 170.0 (ArCH), 124.7 (ArCH), 123.2 (ArCH), 119.7 (ArCH), 117.6 (ArCH), 77.0 (CH₂), 74.9 (CH₂), 71.8 (CH₂), 39.9 (CH₂), 31.8 (CH₂). HR ESIMS calculated for C₇₄H₆₆N₂O₁₀Na⁺=1165.4615; found=1165.4620. Anal. Calcd (%) for C₇₄H₆₆N₂O₁₀: C, 77.74; H, 5.82; N, 2.45; O, 13.99. Found: C, 77.92; H, 5.96, N 2.13.

4.3. Synthesis of calixarene 5a

Oxalyl chloride (0.8 ml, 9.3 mmol) was added dropwise to a solution of 2.14 g 2,3-bis(benzyloxy)benzoic acid (6.4 mmol) in CH₂Cl₂ (50 ml). After adding a drop of DMF, the mixture was stirred until the end of HCl release. After evaporation of solvents and residual oxalyl chloride, the residue was dissolved in 30 ml of dry CH₂Cl₂ and added dropwise to a solution of 2.2 g 5,11,17,23-tetra-*tert*-butyl-25,27-(2aminoethoxy)-26,28-dihydroxy calix[4]arene (3 mmol) and 1.3 ml of triethylamine (9.2 mmol) in 50 ml of dry CH₂Cl₂.

After 18 h under stirring, the mixture was washed with HCl 1 N (100 ml), water (2×50 ml), brine (100 ml), then dried over MgSO₄ and evaporated to dryness. The residue was purified by silica gel column chromatography (EtOAc/cyclohexane 1:2) to give 5a (2.34 g, 57%) as white foam. ¹H NMR (CDCl₃, 300 MHz, 25 °C) δ (ppm): 8.20 (br s, 2H, NH), 7.59 (m, 2H, Ar-H), 7.30–7.41 (m, 20H, Ar-H), 7.15 (s, 2H, Ar-H), 7.03-7.08 (m, 8H, Ar-H), 6.78 (s, 4H, Ar-H subst. rings), 5.13 (s, 4H, O-CH₂-Ar), 5.04 (s, 4H, O-CH₂-Ar), 4.18 $(AB d, 4H, {}^{2}J(H,H)=13.0 Hz, Ar-CH_{2}-Ar ax), 3.93 (t, 4H, J=4.9 Hz, O-$ CH₂-CH₂-NH), 3.65-3.71 (m, 4H, O-CH₂-CH₂-NH), 3.22 (AB d, 4H, 2 J(H,H)=13.0 Hz, Ar–CH₂–Ar eq), 1.31 (s, 18H, (CH₃)₃C unsubst. rings), 0.98 (s, 18H, (CH₃)₃C subst. rings). ¹³C NMR (CDCl₃, 75 MHz, 25 °C) δ (ppm): 166.5 (C=O), 150.2 (ArC), 150.7 (ArC), 149.9 (ArC), 147.4 (ArC), 142.0 (ArC), 137.2 (ArC), 137.1 (ArC), 132.8 (ArC), 129.4 (ArCH), 129.0 (ArCH), 128.9 (ArCH), 128.5 (ArCH), 128.1 (ArCH), 126.0 (ArCH), 125.6 (ArCH), 124.6 (ArCH), 123.4 (ArCH), 117.9 (ArCH), 76.8 (CH₂), 75.0 (CH₂), 72.0 (CH₂), 39.9 (CH₂), 34.4 (Cq), 34.2 (Cq), 32.2 (CH₂), 32.1 (CH₃), 31.5 (CH₃). HR ESIMS calculated for C₉₀H₉₈N₂O₁₀Na⁺=1389.7119; found=1389.7122. Anal. Calcd (%) for $C_{90}H_{98}N_2O_{10};$ C, 79.03; H, 7.22; N, 2.05; O, 11.70. Found: C, 78.59; H, 7.49, N 1.88.

4.4. Synthesis of calixarene 4b

Oxalyl chloride (0.35 ml, 4.08 mmol) was added dropwise to a solution of 0.54 g 1-(benzyloxy)-2-oxo-1.2-dihydropyridine-6carboxylic acid (2.2 mmol) in CH₂Cl₂ (20 ml). After adding a drop of DMF, the mixture was stirred until the end of HCl release. After evaporation of solvents and residual oxalyl chloride, the residue was dissolved in 20 ml of dry CH₂Cl₂ and added dropwise to a solution of 0.55 g 25,27-(2-aminoethoxy)-26,28-dihydroxycalix[4]arene (1.07 mmol) and 0.5 ml of triethylamine (3.6 mmol) in 30 ml of dry CH₂Cl₂. After 18 h under stirring, the mixture was washed with HCl 1 N (20 ml), water $(2 \times 50 \text{ ml})$, brine (100 ml), then dried over MgSO₄ and evaporated to dryness. The residue was purified by silica gel column chromatography (EtOAc) to give 4b (0.625 g, Y=60%) as white powder. ¹H NMR (CDCl₃, 300 MHz, 25 °C) δ (ppm): 8.50 (br s, 2H, NH), 7.76 (s, 2H, Ar-OH), 7.52 (m, 4H, Ar-H), 7.35 (m, 6H, Ar-H), 6.82-6.99 (m, 10H, Ar-H), 6.73-6.80 (m, 4H, Ar-H), 6.49 (m, 2H, Ar-H), 6.19 (m, 2H, Ar-H), 5.32 (s, 4H, O-CH₂-Ar), 4.06 (m, 8H, O-CH₂-CH₂-NH and Ar-CH₂-Ar ax), 3.73 (m, 4H, O-CH₂-CH₂-NH₂), 3.29 (AB d, 4H, ²J(H,H)=13.4 Hz, Ar-CH₂-Ar eq). ¹³C NMR (CDCl₃, 75 MHz, 25 °C) δ (ppm): 161.3 (C=O), 158.9 (C=O), 152.4 (ArC), 150.1 (ArC), 143.7 (ArC), 138.7 (ArC), 138.1 (ArCH), 133.8 (ArC), 133.2 (ArC), 130.7 (ArCH), 129.8 (ArCH), 129.7 (ArCH), 129.2 (ArCH), 129.0 (ArCH), 128.9 (ArCH), 128.0 (ArC), 126.4 (ArCH), 123.8 (ArCH), 123.2 (ArCH), 120.4 (ArCH), 105.1 (ArCH), 79.8 (CH₂), 74.9 (CH₂), 39.9 (CH_2) , 31.7 (CH_2) . HR ESIMS calculated for $C_{58}H_{52}N_4O_{10}Na^+=987.3581$; found=987.3585. Anal. Cald (%) for C₅₈H₅₂N₄O₁₀: C, 72.18; H, 5.43; N, 5.81; O, 16.58. Found: C, 72.05; H, 5.67; N, 5.70.

4.5. Synthesis of calixarene 5b

Oxalyl chloride (0.7 ml, 8.15 mmol) was added dropwise to a solution of 1.44 g 1-(benzyloxy)-2-oxo-1,2-dihydropyridine-6-carboxylic acid (5.87 mmol) in CH₂Cl₂ (50 ml). After adding a drop of DMF, the mixture was stirred until the end of HCl release. After evaporation of solvents and residual oxalyl chloride, the residue was dissolved in 30 ml of dry CH₂Cl₂ and added dropwise to a solution of 2.1 g 5,11,17,23-tetra-*tert*-butyl-25,27-(2-aminoethoxy)-26,28-dihydroxycalix[4]arene (2.85 mmol) and 1 ml of triethylamine (7.2 mmol) in 50 ml of dry CH₂Cl₂. After 18 h under stirring, the mixture was washed with HCl 1 N (100 ml), water (2×50 ml), brine (100 ml), then dried over MgSO₄ and evaporated to dryness. The

residue was purified by silica gel column chromatography (EtOAc/ cyclohexane 3:1) to give **5b** (2.27 g, 63%) as white foam. ¹H NMR (CDCl₃, 300 MHz, 25 °C) δ (ppm): 8.64 (br s, 2H, NH), 7.52 (m, 4H, Ar-H), 7.35 (m, 6H, Ar-H), 6.96 (m, 6H, Ar-H), 6.79 (s, 4H, Ar-H), 6.48 (m, 2H, Ar-H), 6.21 (m, 2H, Ar-H), 5.34 (s, 4H, O-CH₂-Ar), 4.26 (m, 8H, O-CH₂-CH₂-NH and Ar-CH₂-Ar ax), 3.70 (m, 4H, O-CH₂-CH₂-NH₂), 3.28 (AB d, 4H, ²J(H,H)=13.2 Hz, Ar-CH₂-Ar eq), 1.25 (s, 18H, (CH₃)₃C unsubst. rings), 0.98 (s, 18H, (CH₃)₃C subst. rings). ¹³C NMR (CDCl₃, 75 MHz, 25 °C) δ (ppm): 161.4 (C=O), 158.9 (C=O), 149.7 (ArC), 149.0 (ArC), 148.1 (ArC), 143.7 (ArC), 143.0 (ArC), 138.1 (ArCH), 134.0 (ArC), 132.7 (ArC), 130.7 (ArCH), 129.8 (ArCH), 129.0 (ArCH), 127.7 (ArC), 126.3 (ArCH), 125.8 (ArCH), 123.7 (ArCH), 105.7 (ArCH), 80.0 (CH₂), 74.8 (CH₂), 40.0 (CH₂), 34.4 (Cq), 34.2 (Cq), 32.2 (CH₂), 32.0 (CH₃), 31.4 (CH₃). HR ESIMS calculated for $C_{74}H_{84}N_4O_{10}Na^+=1211.6085$; found=1211.6084. Anal. Calcd (%) for C₇₄H₈₄N₄O₁₀: C, 74.72; H, 7.12; N, 4.71; O, 13.45. Found: C, 74.76; H, 7.25; N, 4.54.

4.6. Synthesis of 1,3-CalixCAM 6a

25,27-(2-(2,3-Bis(benzyloxy)-1-carboxamido)ethyl)-26,28-dihy droxycalix[4]arene (0.47 g, 0.41 mmol) and 70 mg of Pd/C (5%) in 20 ml of THF was stirred under 1 atm of H₂. After 96 h, the mixture was filtered on Celite, evaporated to dryness to give 6a (315 mg, 97%) as grey foam. ¹H NMR (CDCl₃, 300 MHz, 25 °C) δ (ppm): 12.53 (br s, Ar-OH), 8.31 (br s, 2H, NH), 8.23 (br s, 2H, Ar-OH), 7.19 (m, 2H, Ar-H), 6.68-7.04 (m, 14H, Ar-H), 6.38 (m, 2H, Ar-H), 4.01-4.12 (m, 8H, O-CH2-CH2-NH and Ar-CH2-Ar ax), 3.36-3.43 (m, 8H, O-CH2- CH_2 -NH₂ and Ar- CH_2 -Ar eq). ¹³C NMR (CDCl₃, 75 MHz, 25 °C) δ (ppm): 171.0 (C=O), 152.2 (ArC), 150.5 (ArC), 146.5 (ArC), 133.5 (ArC), 129.9 (ArCH), 129.5 (ArCH), 128.2 (ArC), 127.0 (ArCH), 125.9 (ArCH), 121.1 (ArCH), 119.1 (ArCH), 118.6 (ArCH), 117.4 (ArCH), 114.5 (ArC), 75.0 (CH₂), 39.9 (CH₂), 32.1 (CH₂). HR ESIMS calculated for C₄₆H₄₂N₂O₁₀Na⁺=805.2737; found=805.2736. Anal. Calcd (%) for C₄₆H₄₂N₂O₁₀·4^{*}H₂O: C, 64.63; H, 5.90; N, 3.28; O, 26.20. Found: C, 64.92; H, 5.83, N 3.05.

4.7. Synthesis of 1,3-CalixHOPO 6b

25,27-(2-(1-Benzyloxy-2-oxo-1,2-dihydropyridine-6- carboxami do)ethyl)-26,28-dihydroxycalix[4]arene (0.85 mg, 0.888 mmol) was added in 30 ml 32% HCl in 60 ml acetic acid. After 96 h under stirring, the mixture was evaporated, 100 ml were added. The mixture was washed with CH₂Cl₂ (3×50 ml). The combined organic layers were washed with water (3×50 ml), brine (50 ml), then dried over MgSO₄ and evaporated to dryness to give **6b** (603 mg, 87%) as yellow foam. ¹H NMR (DMSO-*d*₆, 300 MHz, 25 °C) δ (ppm): 9.13 (br s, 2H, Ar–OH), 8.06 (s, 2H, NH), 7.26 (m, 2H, Ar–H), 7.11 (d, *J*=7.5 Hz, 4H, Ar–H), 6.77 (m, 2H, Ar–H), 6.48–6.56 (m, 4H, Ar–H), 6.42 (d, *J*=6.9 Hz, 2H, Ar–H), 4.17 (AB d, 4H, ²*J*(H,H)=13.0 Hz, Ar–CH₂–Ar ax), 4.06 (m, 4H, O–CH₂–CH₂–NH), 3.80 (d, *J*=5.4 Hz, 4H O–CH₂–CH₂–NH), 3.25 (AB d, 4H, ²*J*(H,H)=13.0 Hz, Ar–CH₂–Ar eq).

¹³C NMR (CDCl₃, 75 MHz, 25 °C) δ (ppm): 160.6 (C=O), 158.0 (C=O), 153.0 (ArC), 151.5 (ArC), 138.5 (ArC), 135.4 (ArCH), 133.2 (ArC), 129.6 (ArCH), 129.1 (ArCH), 128.3 (ArC), 126.1 (ArCH), 120.0 (ArCH), 117.9 (ArCH), 111.8 (ArCH), 74.8 (CH₂), 40.4 (CH₂), 31.7 (CH₂). HR ESIMS calculated for C₄₄H₄₀N₄O₁₀Na⁺=807.2642; found= 807.2644. Anal. Calcd (%) for C₄₄H₄₀N₄O₁₀: C, 67.34; H, 5.14; N, 7.14; O, 20.39. Found: C, 67.38; H, 5.38; N, 6.85.

4.8. Synthesis of 1,3-CalixCAM 7a

5,11,17,23-Tetra-*tert*-butyl-25,27-(2-(2,3-bis(benzyloxy)-1- carb oxamido)ethyl)-26,28-dihydroxycalix[4]arene (1.027 g, 0.75 mmol) and 200 mg of Pd/C (5%) in 25 ml of THF was stirred under 1 atm of H₂. After 96 h, the mixture was filtered on Celite, evaporated to

dryness to give **7a** (735 mg, 98%) as grey foam. ¹H NMR (CDCl₃, 300 MHz, 25 °C) δ (ppm): 8.31 (br s, 2H, NH), 7.26 (m, 2H, Ar–H), 6.87–7.03 (m, 10H, Ar–H), 6.41 (m, 2H, Ar–H), 4.09 (AB d, 4H, ²J(H,H)=12.8 Hz, Ar–CH₂–Ar ax), 3.97 (m, 4H, O–CH₂–CH₂–NH), 3.59 (m, 4H, O–CH₂–CH₂–NH₂), 3.36 (AB d, 4H, ²J(H,H)=12.8 Hz, Ar–CH₂–Ar eq), 1.17 (s, 18H, (CH₃)₃C unsubst. rings), 1.01 (s, 18H, (CH₃)₃C subst. rings). ¹³C NMR (CDCl₃, 75 MHz, 25 °C) δ (ppm): 171.1 (C=O), 150.1 (ArC), 149.7 (ArC), 148.8 (ArC), 146.5 (ArC), 143.6 (ArC), 136.3 (ArC), 133.3 (ArC), 129.4 (ArCH), 128.6 (ArCH), 127.9 (ArC), 126.7 (ArCH), 126.3 (ArCH), 126.0 (ArCH), 118.8 (ArCH), 117.7 (ArCH), 114.5 (ArC), 75.0 (CH₂), 39.9 (CH₂), 34.7 (Cq), 34.4 (Cq), 32.8 (CH₂), 32.2 (CH₃), 31.6 (CH₃). HR ESIMS calculated for C₆₂H₇₄N₂O₁₀ k⁺= 1029.5241; found=1029.5242. Anal. Calcd (%) for C₆₂H₇₄N₂O₁₀: C, 73.93; H, 7.41; N, 2.78; O, 15.88. Found: C, 73.92; H, 7.66, N 2.54.

4.9. Synthesis of 1,3-CalixHOPO 7b

5,11,17,23-Tetra-tert-butyl-25,27-(2-(1-benzyloxy-2-oxo-1,2-dih ydropyridine-6-carboxamido)ethyl)-26,28-dihydroxycalix[4]arene (884 mg, 0.876 mmol) was added in 25 ml 32% HCl in 65 ml acetic acid. After a week under stirring, the mixture was evaporated, 100 ml were added. The mixture was washed with CH₂Cl₂ (3×50 ml). The combined organic layers were washed with water (3×50 ml), brine (50 ml), then dried over MgSO₄ and evaporated to dryness to give **7b** (617 mg, 70%) as orange powder. ¹H NMR (CDCl₃, 300 MHz, 25 °C) δ (ppm): 6.89–7.23 (m, 3H, ArH), 7.01 (s, 4H, Ar-H), 6.75 (s, 4H, Ar-H), 4.06-4.21 (m, 12H, O-CH₂-CH₂-NH and Ar-CH₂-Ar ax), 3.29 (AB d, 4H, ²/(H,H)=13.2 Hz, Ar-CH₂-Ar eq), 1.28 (s, 18H, (CH₃)₃C unsubst. rings), 0.93 (s, 18H, (CH₃)₃C subst. rings). ¹³C NMR (CDCl₃, 75 MHz, 25 °C) δ (ppm): 160.2 (C=O), 157.3 (C=O), 150.2 (ArC), 149.6 (ArC), 147.9 (ArC), 142.6 (ArC), 135.1 (ArCH), 132.7 (ArC), 128.2 (ArC), 126.2 (ArCH), 126.1 (ArCH), 125.6 (ArCH), 117.3 (ArCH), 112.2 (ArCH), 74.5 (CH₂), 40.4 (CH₂), 34.3 (Cq), 34.2 (Cq), 32.1 (CH₂), 32.0 (CH₃), 31.4 (CH₃). HR ESIMS calculated for C₆₀H₇₂N₄O₁₀ $Na^+=1031.5146$; found=1031.5145. Anal. Calcd (%) for $C_{60}H_{72}N_4O_{10}$: C, 71.40; H, 7.19; N, 5.55; O, 15.85. Found: C, 71.61; H, 7.32; N, 5.21.

4.10. Synthesis of 1,3-CalixCAMS 8a

25,27-(2-(N-(2,3-Bis(hydroxy)-1-benzamido))ethyl)-26,28-dihy droxycalix[4]arene (106 mg, 0.135 mmol) in H₂SO₄ (2 ml) was stirred at 50 °C for 16 h. The mixture was then allowed to cool to room temperature, poured in Et₂O (50 ml). The resulting precipitate was filtered off under argon and dried under vacuum to give **8a** as a hygroscopic brown powder (140 mg, 82%). ¹H NMR (D₂O, 300 MHz, 25 °C) δ (ppm): 7.26 (d, *J*=1.9 Hz, 2H, Ar-*H*), 7.49 (s, 4H, Ar-H), 7.24 (s, 4H, Ar-H), 7.14 (d, J=1.9 Hz, 2H, Ar-H), 4.23 (t, J=4.2 Hz, 4H, O-CH₂-CH₂-N), 4.13 (t, J=4.2 Hz, 4H, O-CH₂-CH₂-N), 4.05 (AB d, 4H, ²J(H,H)=13.8 Hz, Ar-CH₂-Ar ax), 3.48 (AB d, 4H, $^{2}J(H,H)=13.8$ Hz, Ar-CH₂-Ar eq). ^{13}C NMR (D₂O, 75 MHz, 25 °C) δ (ppm): 169.8 (C=O), 155.2 (ArC), 153.9 (ArC), 148.8 (ArC), 145.1 (ArC), 139.8 (ArC), 134.2 (ArC), 133.7 (ArC), 133.5 (ArC), 127.4 (ArC), 127.0 (ArCH), 126.9 (ArCH), 126.7 (ArCH), 117.8 (ArC), 117.2 (ArCH), 115.3 (ArCH), 74.4 (CH₂), 40.1 (CH₂), 31.0 (CH₂). HR ESIMS calculated for C₄₆H₄₂N₂O₂₈S₆Na⁺=1285.0141; found=1285.0136. Anal. Calcd (%) for $C_{46}H_{42}N_2O_{28}S_6 \cdot 6^*H_2O$: C, 40.29; H, 3.97; N, 2.04; O, 39.67; S, 14.03. Found: C, 39.98; H, 4.12; N, 1.79.

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References and notes

- 1. Hamilton, J. G. Rev. Mod. Phys. 1948, 20, 718-728.
- 2. Bulman, R. A. Coord. Chem. Rev. 1980, 31, 221-250.
- Galle, P. Uranium. In *Toxiques Nucléaires*; Masson: Paris, 1997; Chapter XIII, pp 185–205.
- (a) Sawicki, M.; Siaugue, J.-M.; Jacopin, C.; Moulin, C.; Bailly, T.; Burgada, R.; Meunier, S.; Baret, P.; Pierre, J.-L.; Taran, F. *Chem.—Eur. J.* 2005, *11*, 3689–3697; (b) Bailly, T.; Burgada, R.; Prange, T.; Lecouvey, M. *Tetrahedron Lett.* 2002, *44*, 189–192; (c) Burgada, R.; Bailly, T.; Prange, T.; Lecouvey, M. *Tetrahedron Lett.* 2007, *48*, 2315– 2319; (d) Xu, G.; Yang, C.; Liu, B.; Wu, X.; Xie, Y. *Heteroat. Chem.* 2004, *15*, 251–257.
- 5. (a) Martinez, A. B.; Cabrini, R. L.; Ubios, A. M. Health Phys. 2000, 78, 668–671;
 (b) Ubios, A. M.; Braun, E. M.; Cabrini, R. L. Health Phys. 1998, 75, 610–613; (c)
 Ubios, A. M.; Braun, E. M.; Cabrini, R. L. Health Phys. 1994, 66, 540–544; (d)
 Fukuda, S.; lida, H.; Ikeda, M.; Yan, X.; Xie, Y. Health Phys. 2005, 89, 81–88.
- 6. Durbin, P.; Kullgren, B.; Ebbe, S. N.; Xu, J.; Raymond, K. N. Health Phys. 2000, 78, 511–521.
- (a) Sylwester, E. R.; Allen, P. G.; Dharmawardana, U. R.; Sutton, M. Inorg. Chem. 2001, 40, 2835–2841; (b) Domingo, J. L.; Llobet, J. M.; Corbella, J. Fundam. Appl. Toxicol. 1990, 14, 88–95.
- (a) Ramounet-Le Gall, B.; Grillon, G.; Rateau, G.; Burgada, R.; Bailly, T.; Fritsch, P. Radiat. Prot. Dosim. 2003, 105, 535–538; (b) Gorden, A. E. V.; Xu, J.; Raymond, K. N.; Durbin, P. Chem. Rev. 2003, 103, 4207–4282.
- Leydier, A.; Lecerclé, D.; Pellet-Rostaing, S.; Favre-Réguillon, A.; Taran, F.; Lemaire, M. *Tetrahedron* 2008, 64, 6662–6669.
- (a) Gutsche, C. D. Calixarenes Revisited. In Monographs in Supramolecular Chemistry; Stoddart, J. F., Ed.; The Royal Society: Cambridge, 1998; Vol. 6; (b) Asfari, Z.; Böhmer, V.; Harrowfield, J.; Vicens, J. Calixarenes 2001; Kluwer Academic: Dordrecht, 2001; (c) McKervey, M. A.; Arnaud-Neu, F.; Schwing-Weill, M.-J. In Comprehensive Supramolecular Chemistry; Gokel, G., Ed.; Pergamon: Oxford, UK, 1996; Vol. 1, pp 537–603; (d) Wieser, C.; Dieleman, C. B.; Matt, D. Coord. Chem. Rev. 1997, 165, 93–161; (e) Ikeda, A.; Shinkaï, S. Chem. Rev. 1997, 97, 1713–1734; (f) Harvey, P. D. Coord. Chem. Rev. 2002, 233–234, 289–309; (g) Sliwa, W. J. Inclusion Phenom. Macrocycl. Chem. 2005, 52, 13–37.
- (a) In Calixarenes: A Versatile Class of Macrocyclic Compounds; Vicens, J., Bohmer, V., Eds.; Kluwer Academic: Dordrecht, Netherland, 1991; (b) Rebek, J. Chem.

Commun. 2000, 637–643; (c) Le Gac, S.; Luhmer, M.; Reinaud, O.; Jabin, I. *Tet-rahedron* 2007, 63, 10721–10730; (d) Zhao, B.-T.; Blesa, M.-J.; Le Derf, F.; Canevet, D.; Benhaoua, C.; Mazari, M.; Allain, M.; Salle, M. *Tetrahedron* 2007, 63, 10768–10777.

- (a) Sonoda, M.; Nishida, M.; Ishii, D.; Yoshida, I. Anal. Sci. 1999, 15, 1207–1213;
 (b) Araki, K.; Hashimoto, N.; Otsuka, H.; Nagasaki, T.; Shinkai, S. Chem. Lett. 1993, 829–832.
- Shinkai, S.; Koreishi, H.; Ueda, K.; Arimura, T.; Manabe, O. J. Am. Chem. Soc. 1987, 109. 6371–6375.
- Beer, P. D.; Brindley, G. D.; Danny Fox, O.; Grieve, A.; Ogden, M. I.; Szemes, F.; Drew, M. G. D. J. Chem. Soc., Dalton Trans. 2002, 3101–3111.
- Dasaradhi, L.; Stark, P. C.; Huber, V. J.; Smith, P. H.; Jarvinen, G. D.; Gopalan, A. S. J. Chem. Soc., Perkin Trans. 2 1997, 1187–1192.
- Barboso, S.; Carrera, A. G.; Matthews, S. E.; Arnaud-Neu, F.; Bohmer, V.; Dozol, J.-F.; Rouquette, H.; Schwing-Weill, M.-J. J. Chem. Soc., Perkin Trans. 2 1999, 719–724.
- 17. Lambert, T. N.; Dasaradhi, L.; Huber, V. J.; Gopalan, A. S. J. Org. Chem. **1999**, 64, 6097–6101.
- (a) Xu, J.; Durbin, P. W.; Kullgren, B.; Ebbe, S. N.; Uhlir, L. C.; Raymond, K. N. J. Med. Chem. 2002, 45, 3963–3971; (b) Burgada, R.; Bailly, T.; Noel, J. P.; Gomis, J. M.; Valleix, A.; Ansoborlo, E.; Henge-Napoli, M. H.; Paquet, F.; Gourmelon, P. J. Labelled Compd. Radiopharm. 2001, 44, 13–19.
- (a) Laursen, B.; Denieul, M.-P.; Skrydstrup, T. *Tetrahedron* **2002**, *58*, 2231–2238;
 (b) Gardner, R. A.; Kinkade, R.; Wang, C.; Phanstiel, O. I. V. J. Org. Chem. **2004**, *69*, 3530–3537.
- (a) Halouani, H.; Dumazet-Bonnamour, I.; Perrin, M.; Lamartine, R. J. Org. Chem. 2004, 69, 6521–6527; (b) Collins, E. M.; McKervey, M. A.; Madigan, E.; Moran, M. B.; Owens, M.; Ferguson, G.; Harris, S. J. Chem. Soc., Perkin Trans. 1 1991, 3137–3142; (c) Casnati, A.; Massera, C.; Pelizzi, N.; Stibor, I.; Pinkassik, E.; Ugozzoli, F.; Ungaro, R. Tetrahedron Lett. 2002, 43, 7311–7314; (d) Danila, C.; Bolte, M.; Boehmer, V. Org. Biomol. Chem. 2005, 3, 172–184.
- 21. Jaime, C.; Mendoza, J.; Prados, P.; Nieto, P. M.; Sanchez, C. J. Org. Chem. **1991**, *56*, 3372–3376.
- Thomas, F.; Beguin, C.; Pierre, J. L.; Serratrice, G. Inorg. Chim. Acta 1999, 291, 148–157.